



Review of Environmental Culturing – [REDACTED]

Collection Date: [REDACTED]

For purposes of evaluating cleanliness, Evaclean uses the Mulvey¹ findings for determining a benchmark. The current quantitative standards for aerobic colony counts (ACC) on surfaces/hand touch sites should not exceed 2.5 cfu/cm². The qualitative standard states that any pathogen isolated should be <1 cfu/cm².

Samples were taken without cleaning from two surfaces²:

- a. Table
- b. TV Remote Control

Both surfaces were then sprayed with PURTABS, NaDCC, at a concentration of 4306 ppm. After the disinfectant was allowed to air dry, culture samples were taken of both surfaces.

Surface	Pre-Clean Swab	CFU/Growth	Post-Spray Swab	CFU
Table	Staphylococcus epidermis Pantoea species	1 1	No growth	0
Remote	Staphylococcus epidermis	In thio Broth	No growth	0

Disinfecting the surfaces with PURTABS resulted in no growth of bacteria.

¹ D. Mulvey, et. al., Finding a Benchmark for Monitoring Hospital Cleanliness, *Journal of Hospital Infection* 77 (2011) 25-30.

² See "Final Report" CD Laboratories

BACTERIAL FINDINGS

Pantoea Species

Pantoea agglomerans (formerly *Enterobacter agglomerans*) is a motile peritrichous, non-sporeforming, Gram-negative aerobic bacilli in the *Enterobacteriaceae* family (1, 2). It is commonly found in the ecological niches such as water, soil, sewage, seeds, vegetables, feculent material and foodstuffs, as well as reported as both commercial and opportunistic pathogens of animals and humans (3, 4). This opportunistic pathogen isolated from clinical specimens including blood, wounds, urine, throat, and internal organs (5).

P. agglomerans is recognized as a plant pathogen. In the mid-1960s it was identified in nosocomial infections (6, 7). *P. agglomerans* is the most frequent species associated with human infections (1, 8). Hospital outbreaks due to contamination of anesthetic agent propofol, blood products, parenteral nutrition, and transference tubes used for intravenous hydration have been demonstrated (8, 9, 10).

P. agglomerans has been implicated in pneumonia, wound infections, septicemia, bacteremia, urinary tract infection, meningitis, lung and brain abscess, septicemia, osteomyelitis, septic arthritis, peritonitis and coledithiasis. The organism is generally regarded as opportunistic, of low virulence, low degree of toxicity and with little intrinsic invasiveness but can cause infection even in the healthy individuals with immunocompetent system (11, 12).

P. agglomerans is causative agent of infection in children and elderly persons.

REFERENCES:

1. Cruz AT, Cazacu AC, Allen CH. *Pantoea agglomerans*, a plant pathogen causing human disease. *J Clin Microbiol.* 2007;45:1989–1992. [PMC free article] [PubMed] [Google Scholar]
2. Sharma M, Dogra BB, Misra R, Gandham N, Sardar M, Jadhav S. Multidrug resistant *Pantoea agglomerans* in a patient with septic arthritis—a rare report from India. *Int J Microbiol Res.* 2012;4:263–265. [Google Scholar]
3. Eickhoff TC, Steinhauer BW, Finland M. The Klebsiella-Enterobacter-Serratia division. Biochemical and serologic characteristics and susceptibility to antibiotics. *Ann Intern Med.* 1966;65:1163–1179. [PubMed] [Google Scholar]
4. Liberto MC, Matera G, Puccio R, Lo Russo T, Colosimo E, Focà E. Six cases of sepsis caused by *Pantoea agglomerans* in a teaching hospital. *New Microbiol.* 2009;32:119–123. [PubMed] [Google Scholar]
5. Gavini F, Mergaert J, Bej A, Mielcarek C, Izard D, Kersters K, et al. Transfer of *Enterobacter agglomerans* (Beijerinck 1888) Ewing and Fife 1972 to *Pantoea* gen. nov. as *Pantoea agglomerans* comb. nov. and Description of *Pantoea dispersa* sp. nov. *Int J Syst Bacteriol.* 1989;39:337–345. [Google Scholar]
6. Steinhauer BW, Eickhoff TC, Kislak JW, Finland M. The Klebsiella-Enterobacter-Serratia division. Clinical and epidemiologic characteristics. *Ann Intern Med.* 1966;65:1180–1194. [PubMed] [Google Scholar]
7. Starr MP, Chatterjee AK. The genus *Erwinia*: enterobacteria pathogenic to plants and animals. *Annu Rev Microbiol.* 1972;26:389–426. [PubMed] [Google Scholar]
8. Boszczowski I, Nóbrega de Almeida Júnior J, Peixoto de Miranda EJ, Pinheiro Freire M, Guimarães T, Chaves CE, et al. Nosocomial outbreak of *Pantoea agglomerans* bacteraemia associated with contaminated anticoagulant citrate dextrose solution: new name, old bug? *J Hosp Infect.* 2012;80:255–258. [PubMed] [Google Scholar]
9. Alvarez FE, Rogge KJ, Tarrand J, Lichtiger B. Bacterial contamination of cellular blood components. A retrospective review at a large cancer center. *Ann Clin Lab Sci.* 1995;25:283–290. [PubMed] [Google Scholar]
10. Habsah H, Zeehaida M, Van Rostenberg H, Noraida R, Wan Pauzi WI, Fatimah I, et al. An outbreak of *Pantoea* spp. in a neonatal intensive care unit secondary to contaminated parenteral nutrition. *J Hosp Infect.* 2005;61:213–218. [PubMed] [Google Scholar]
11. Rosenfeld R, Spiegelblatt L, Chicoine R, Laverdiere M. Thorn-induced periostitis associated with *Enterobacter agglomerans* infection. *Can Med Assoc J.* 1978;119:925–928. [PMC free article] [PubMed] [Google Scholar]
12. Al-Damluji S, Dickinson CM, Beck A. *Enterobacter agglomerans*: a new cause of primary pneumonia. *Thorax.* 1982;37:865–866. [PMC free article] [PubMed] [Google Scholar]

Staphylococcus epidermidis

Whereas previously only regarded as an innocuous commensal microorganism on the human skin, *Staphylococcus epidermidis* is nowadays seen as an important opportunistic pathogen. It is now the most frequent cause of nosocomial infections, at a rate about as high as that due to its more virulent cousin *Staphylococcus aureus*¹. In particular, *S. epidermidis* represents the most common source of infections on indwelling medical devices. This likely stems from the fact that *S. epidermidis* is a permanent and ubiquitous colonizer of human skin, and the resulting high probability of device contamination during insertion². While *S. epidermidis* infections only rarely develop into life-threatening diseases, their frequency and the fact that they are extremely difficult to treat represent a serious burden for the public health system. The costs related to vascular catheter-related bloodstream infections caused by *S. epidermidis* amount to an estimated \$ 2 billion annually in the United States alone³⁻⁵. Treatment is complicated by specific antibiotic resistance genes and the formation of biofilms, multicellular agglomerations that have intrinsic resistance to antibiotics and mechanisms of host defense³. Furthermore, recent investigation has identified specific molecular determinants facilitating *S. epidermidis* immune evasion and ability to cause chronic disease.

Staphylococci are common bacterial colonizers of the skin and mucous membranes of humans and other mammals⁴. *S. epidermidis* in particular is the most frequently isolated species from human epithelia. It colonizes predominantly the axillae, head, and nares⁵.

S. epidermidis belongs to the group of coagulase-negative staphylococci (CoNS), which is distinguished from coagulase-positive staphylococci such as *S. aureus* by lacking the enzyme coagulase. The species shows a high degree of diversity with 74 identified sequence types (STs)⁶.

Among CoNS, *S. epidermidis* clearly causes the greatest number of infections^{2,9}. In clinical microbiology, CoNS are often not further specified, as the major interest is in making a distinction between *S. aureus* and other staphylococci. However, based on reports that have performed species identification^{1,5}, one can assume that the vast majority of non-specified CoNS infections are due to *S. epidermidis*. Particularly, *S. epidermidis* represents the most frequent causative agent involved with infections of any type of indwelling medical devices, such as peripheral or central intravenous catheters (CVCs)⁹. These infections usually commence with the introduction of bacteria from the skin of the patient or that of health care personnel during device insertion and have increased in number most likely owing to the increased use of such devices^{1,18}. *S. epidermidis* now accounts for at least 22% of bloodstream infections in intensive care unit patients in the USA, which occur in at least 4–5/1000 CVC insertions^{1,18}.

REFERENCES

1. CDC. National Nosocomial Infections Surveillance (NNIS) System Report, data summary from January 1992 through June 2004, issued October 2004. *Am J Infect Control*. 2004;32:470–485. [[PubMed](#)] [[Google Scholar](#)]
2. Uckay I, et al. Foreign body infections due to *Staphylococcus epidermidis*. *Ann Med*. 2009;41:109–119. [[PubMed](#)] [[Google Scholar](#)]
3. Costerton JW, Stewart PS, Greenberg EP. Bacterial biofilms: a common cause of persistent infections. *Science*. 1999;284:1318–1322. [[PubMed](#)] [[Google Scholar](#)]
4. Kloos W, Schleifer KH. In: *Bergey's Manual of Systematic Bacteriology*. PHA S, S M, ME S, JG H, editors. Baltimore: Williams & Wilkins; 1986. [[Google Scholar](#)]
5. Kloos WE, Musselwhite MS. Distribution and persistence of *Staphylococcus* and *Micrococcus* species and other aerobic bacteria on human skin. *Appl Microbiol*. 1975;30:381–385. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

6. Miragaia M, Thomas JC, Couto I, Enright MC, de Lencastre H. Inferring a population structure for *Staphylococcus epidermidis* from multilocus sequence typing data. *J Bacteriol.* 2007;189:2540–2552. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
7. Li M, Wang X, Gao Q, Lu Y. Molecular characterization of *Staphylococcus epidermidis* strains isolated from a teaching hospital in Shanghai, China. *J Med Microbiol.* 2009;58:456–461. [[PubMed](#)] [[Google Scholar](#)]
8. Zhang YQ, et al. Genome-based analysis of virulence genes in a non-biofilm-forming *Staphylococcus epidermidis* strain (ATCC 12228) *Mol Microbiol.* 2003;49:1577–1593. [[PubMed](#)] [[Google Scholar](#)]
9. Rogers KL, Fey PD, Rupp ME. Coagulase-negative staphylococcal infections. *Infect Dis Clin North Am.* 2009;23:73–98. [[PubMed](#)] [[Google Scholar](#)]
10. Chu VH, et al. Coagulase-negative staphylococcal prosthetic valve endocarditis—a contemporary update based on the International Collaboration on Endocarditis: prospective cohort study. *Heart.* 2009;95:570–576. [[PubMed](#)] [[Google Scholar](#)]
11. Massey RC, Horsburgh MJ, Lina G, Hook M, Recker M. The evolution and maintenance of virulence in *Staphylococcus aureus*: a role for host-to-host transmission? *Nat Rev Microbiol.* 2006;4:953–958. [[PubMed](#)] [[Google Scholar](#)]
12. Harder J, Schroder JM. Antimicrobial peptides in human skin. *Chem Immunol Allergy.* 2005;86:22–41. [[PubMed](#)] [[Google Scholar](#)]
13. Faurschou M, Borregaard N. Neutrophil granules and secretory vesicles in inflammation. *Microbes Infect.* 2003;5:1317–1327. [[PubMed](#)] [[Google Scholar](#)]
14. Yao Y, Sturdevant DE, Otto M. Genomewide analysis of gene expression in *Staphylococcus epidermidis* biofilms: insights into the pathophysiology of *S. epidermidis* biofilms and the role of phenol-soluble modulins in formation of biofilms. *J Infect Dis.* 2005;191:289–298. [[PubMed](#)] [[Google Scholar](#)]
15. Khardori N, Yassien M, Wilson K. Tolerance of *Staphylococcus epidermidis* grown from indwelling vascular catheters to antimicrobial agents. *J Ind Microbiol.* 1995;15:148–151. [[PubMed](#)] [[Google Scholar](#)]
16. Duguid IG, Evans E, Brown MR, Gilbert P. Effect of biofilm culture upon the susceptibility of *Staphylococcus epidermidis* to tobramycin. *J Antimicrob Chemother.* 1992;30:803–810. [[PubMed](#)] [[Google Scholar](#)]
17. Duguid IG, Evans E, Brown MR, Gilbert P. Growth-rate-independent killing by ciprofloxacin of biofilm-derived *Staphylococcus epidermidis*: evidence for cell-cycle dependency. *J Antimicrob Chemother.* 1992;30:791–802. [[PubMed](#)] [[Google Scholar](#)]

Conclusion

Cleaning of high touch point surfaces has been insufficient in completely eliminating bacterial counts. Enhanced disinfection using NaDCC demonstrated no bacterial growth after 48 hours. The recommendation is to add enhanced disinfection to daily general cleaning in the common areas.

Disinfection should be done on a daily basis during the night shift using a 4306 PPM concentration of NaDCC. Evaluation of additional surfaces should be conducted on a monthly basis or until CFU p/cm² is achieved.



Patient Name: [REDACTED]
Date of Birth: [REDACTED]
Gender: M
Age: 0D
Unit:
Room/Bed:

Accession #: [REDACTED]
Collection Date: [REDACTED]
Received in Lab: [REDACTED] 11:42
Resulted Date: [REDACTED] 09:00
Ordering Phys.: PROVIDER, UNSPECIFIED
Organization: [REDACTED]

Comments

Test Name	Result	Units	Flag	Ref. Range	Site
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CULTURE, ENVIRONMENTAL

Result

SEE COMMENTS

Site:

Table A

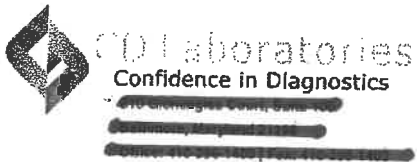
Organism 1: *Staphylococcus epidermidis*

Growth: 1 CFU

Organism 2: *Pantoea species*

Growth: 1 CFU

*** FINAL REPORT ***



FINAL REPORT

Patient Name: ENVIRONMENTAL
 Date of Birth: [REDACTED]
 Gender: M
 Age: 0D
 Unit:
 Room/Bed:

Accession #: [REDACTED]
 Collection Date: [REDACTED]
 Received in Lab: [REDACTED] 11:43
 Resulted Date: [REDACTED] 10:29
 Ordering Phys.: [REDACTED]
 Organization: [REDACTED]

Comments

Test Name	Result	Units	Flag	Ref. Range	Site
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CULTURE, ENVIRONMENTAL

Result

SEE COMMENTS

Site:

Table B

Culture results:

No growth after 48 hours.

*** FINAL REPORT ***



CD Laboratories
Confidence in Diagnostics

Address: [REDACTED]
Phone: [REDACTED]

FINAL REPORT

Patient Name: ENVIRONMENTAL
Date of Birth: [REDACTED]
Gender: [REDACTED]
Age: 0D
Unit:
Room/Bed:

Accession #: 4420042
Collection Date: [REDACTED]
Received in Lab: [REDACTED]
Resulted Date: [REDACTED] 09:00
Ordering Phys.: [REDACTED] UNSPECIFIED
Organization: [REDACTED] CHASE

Comments

Test Name	Result	Units	Flag	Ref. Range	Site
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CULTURE, ENVIRONMENTAL

Result

SEE COMMENTS

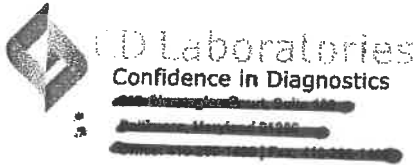
Site:

Remote A

Organism 1: Staphylococcus epidermidis

Growth: In Thio Broth

*** FINAL REPORT ***



FINAL REPORT

Patient Name: [REDACTED]
Date of Birth: [REDACTED]
Gender: [REDACTED]
Age: 0D
Unit:
Room/Bed:

Accession #: [REDACTED]
Collection Date: [REDACTED]
Received in Lab: [REDACTED]
Resulted Date: [REDACTED]
Ordering Phys.: [REDACTED] SPECIFIED
Organization: [REDACTED]

Comments

Test Name	Result	Units	Flag	Ref. Range	Site
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CULTURE, ENVIRONMENTAL

Result

SEE COMMENTS

Site:

Reomte B

Culture results:

No growth after 48 hours.

*** FINAL REPORT ***